

提高现代超级稻产量潜力的栽培生理研究途径探讨

顾骏飞 陈颖 毛倚琦

(扬州大学 江苏省作物遗传生理国家重点实验室培育点/粮食作物现代产业技术创新中心,江苏 扬州 225009;
第一作者:gujf@yzu.edu.cn)

摘要:水稻籽粒灌浆期同化物供应不足会导致籽粒灌浆不充分,从而影响产量,这种现象在大穗型、高库容的现代超级稻品种中尤为明显,直接限制了超级稻品种产量潜力的发挥。灌浆期水稻同化物的供给主要来源于花前同化物的积累与花后同化物的合成,它们均依赖于冠层群体光合能力。因此,加强相关过程的生理生态研究对揭示制约现代超级稻产量潜力的关键性因素具有重要意义。作者综述了相关领域的研究进展,并从叶片光合生理、冠层光氮匹配和根-冠水分平衡等方面,对提高现代超级稻产量潜力的栽培生理途径进行了探讨。同时,作者总结了模型分析在综合栽培生理认识、发现超级稻产量潜力限制性因子及其生理机制等方面的作用。

关键词:光合作用;产量潜力;冠层光氮匹配;根-冠水分平衡

中图分类号:S511.048 文献标识码:A 文章编号:1006-8082(2017)03-0001-05

现代超级稻品种穗型大,穗粒数多,有极高的库容,产量潜力能够达到 $1\,000\text{ kg}/667\text{ m}^2$ 以上^[1]。但是在实际生产中,以江苏为例,超级稻单产只在 $600\sim700\text{ kg}/667\text{ m}^2$ 之间,且表现出灌浆不充分、结实率低,影响了产量潜力的发挥。超级稻品种的结实率往往比常规稻品种低10%以上^[2]。现代超级稻品种灌浆不充分、结实率低,主要原因是灌浆期同化物的供给不充分所造成。要是能突破灌浆期同化物供应的瓶颈,将有望进一步提高我国超级稻的产量。

灌浆期水稻同化物的供给主要来源于两个方面(图1):一是花前累积的同化物,它们来源于花前冠层群体的光合,主要以非结构性碳水化合物的形式贮存在茎鞘中,对产量的贡献为30%左右^[3-4];二是花后冠层群体的光合,其中剑叶光合对产量贡献较大。以下将分别从叶片光合与冠层光合角度来分析提高水稻净同化力的限制性因子。

1 叶片光合作用

光合作用是指绿色植物通过叶绿素吸收光能,同化 CO_2 和 H_2O ,制造有机物并释放 O_2 的过程,根据是否需要光的参与,分为光反应和暗反应两个过程。在光反应过程中,类囊体上的电子传递链捕获和利用光能,合成暗反应需要的能量ATP和还原剂NADPH。光反应形成的ATP和NADPH供给暗反应同化 CO_2 形成碳水化合物。在暗反应中, CO_2 受体为1,5-二磷酸核酮糖(RuBP),在核酮糖二磷酸羧化酶(Rubisco)等一系列酶的作用下, CO_2 被还原成磷酸丙糖3-磷酸甘油醛(3-

GAP)。磷酸丙糖不能直接透过叶绿体内膜,其必需由磷酸运转器与Pi对等交换才能出入叶绿体。当Pi不足或长期光照时,磷酸丙糖会在叶绿体内积累,形成淀粉粒。

因此在低光强下,囊体上的电子传递受限,影响ATP的合成,从而影响RuBP再生(RuBP再生限制);在饱和光强下,暗反应关键酶Rubisco的数量与活性,及反应底物 CO_2 的浓度是限制光合作用的最关键因素(Rubisco羧化限制);在长期光照下,大量形成的光合产物需要借助叶绿体膜上的磷酸运转器及时转运出叶绿体,此时光合速率受细胞质中无机磷运转速率限制(磷酸丙糖运转限制)^[5-6]。根据叶片光合的限制性因子,国内外学者开展了很多研究,希望突破叶片光合限制,提高同化物的供给,例如,通过提高Rubisco活化酶(RCA)的活性来改善Rubisco活性^[7],通过提高Rubisco酶对 CO_2 (相对于 O_2)的专一性(Sc/o)来改善Rubisco活性^[8],降低光呼吸耗损^[9],过表达SBPase基因改善RuBP的再生能力^[10-11],通过增加细胞色素b6f复合体含量提高叶绿体电子传递^[12],引入蓝细胞的 CO_2 浓缩机制来提高反应底物 CO_2 浓度^[13],过表达水通道蛋白NtAQP1

收稿日期:2016-11-20

基金项目:国家重点基础研究发展计划("973"计划)(2015CB150400);国家自然科学基金(31501254);江苏省自然科学基金(BK20140480);中国博士后基金(2014M550312, 2015T80590);江苏省高校自然科学基金(14KJB210007)

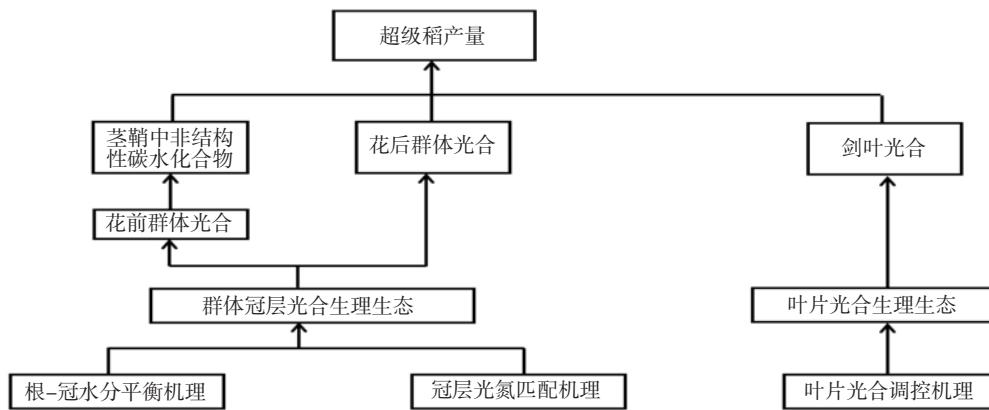


图 1 提高现代超级稻产量潜力的栽培生理途径分解

提高叶肉导度从而提高反应底物 CO_2 浓度^[14-15], 将高效的 C_4 途径引入水稻来消除光合作用的氧抑制^[16]。但是将这些分子生物学成果运用于水稻的大田生产中, 预计需要 15 年甚至更长的时间^[16-17]。针对大田生产中广泛使用的梗型超级稻品种沈农 265 和农家旱稻品种毫格劳, 我们对它们及它们杂交组合后代进行了遗传与生理分析, 发现气孔与叶肉导度是高光强下影响光合最主要的限制性因子^[18-19]。Adachi 等^[20-22]研究发现, 水稻品种 Takanari 干物质积累比水稻品种 Koshihikari 高 20%~30%, 主要是因为 Takanari 根系具有较高的导水率, 能够维持高的叶片水势, 保持气孔开放, 有较高的气孔导度。Yang 等^[23]也发现, 目前大田高氮肥施用条件下氮素利用效率降低是由于水稻根系导水率低, 吸水能力不足, 气孔开放程度低, 气孔导度小所致。因此, 如何通过栽培措施调节根系、根系-叶片水分平衡, 提高气孔导度, 可能是提高水稻光合能力的一个潜在的靶标。

2 冠层光合作用

相对于叶片光合, 冠层群体的光合与水稻干物质的积累、同化物的供应联系更紧密^[24]。冠层结构大大提高了作物光能利用效率。对于水稻等 C_3 作物叶片来说, 当光合有效辐射 (Photosynthetically active radiation, PAR) 达到 700~1 000 $\mu\text{mol}/(\text{m}^2 \cdot \text{s})$ 时, 叶片的光合值达到上限, 光合能力不再随着光强的提高而增加, 称为光饱和点。而当光合有效辐射增加到 2 000 $\mu\text{mol}/(\text{m}^2 \cdot \text{s})$, 冠层光合仍然没有达到饱和, 此时, 冠层光合是叶片光合的 3 倍^[25-26]。这是因为除了冠层顶部的光饱和叶 (如剑叶等), 冠层内阴影部分 70% 的叶片能够吸收占总光能 30% 的散射光与漫射光, 贡献冠层约 50% 的光合能力^[27]。因此, 研究整个冠层的光合生理生态对水稻

的干物质积累非常重要。合理的冠层结构历来受到国内外育种和栽培生理专家的重视与关注^[24,28-29]。相关研究普遍认为, 茎叶夹角小、直立叶片构成的冠层有利于群体受光, 对群体光合作用和物质生产有利。但在水稻高产实践中发现, 基于株型选择的, 具有矮秆、直立叶片等特性的新株型 (New type plant-NTP) 水稻品种的产量往往比基于产量选择的超级稻品种的产量低^[29], 说明人们对群体冠层结构和功能的认识仍然是片面的, 不系统的, 需要深入理解作物高产群体冠层结构的生理生态基础及其与光温等生态因子的匹配机理。

3 提高冠层光合作用的途径

光是绿色植物进行光合作用的能量来源。优良的冠层结构有利于光在群体中的分布, 有利于群体受光。除了光的分布, 氮素作为 Rubisco 酶与叶绿体的重要组成部分, 它在冠层中的分布也是限制植物体内物质合成的关键因素^[30-32]。和光一样, 氮素在冠层内的分布也呈现顶部高、基部低的梯度分布。这种氮素的梯度分布相对于均匀分布, 冠层光合提高了 20%^[33], 这种现象被认为是植物对外界光环境的适应, 是为了提高冠层光合同化力和氮素利用效率^[33-35]。但是通常冠层中氮素的分布并不是最优的^[36-38], Hikosaka 等^[39]通过研究发现, 如果能够调控冠层中的氮素分布, 使之与光的分布一致, 达到最优, 冠层光合能额外再提高 20%。Dingkuhn 等^[40]在 1991 年就提出, 优化冠层氮的分布 (即加大氮素的分布梯度差, 增加上部叶片含氮量, 减少下部叶片含氮量) 应该作为育种中的重要选择靶标。但是, 氮素在冠层中的分布调控机理非常复杂, 除了光强外, 也受其他因素调控, 如环境中红光/远红光比例^[41]、植株体内激素^[42]、叶龄^[32]、库源关系^[31]等。在栽培调控措施方面, Hikosaka 等^[34,43-44]的研究表明, 高施氮水平能够改善氮

素与光的梯度分布,但是受发育时期影响^[45]。而 Sinclair 等^[46–47]却持不同观点,认为氮肥水平对光、氮的分布没有影响。因此,在生产中栽培措施能否优化光、氮分布,充分利用植株体内氮素与光合效率空间梯度特点,实现作物群体光合效率的最大化,值得深入探讨。鉴于冠层光合的复杂性,冠层中的光、氮分布及其对冠层光合的贡献研究常常借助于模型模拟分析^[27,39,48–50]。同时,植物冠层群体是动态变化的,苗期叶片的快速生长与冠层结构的建成,有利于光的有效截获和花前干物质的积累^[40,51]。开花后作物氮素吸收、分配与植株体内氮素代谢影响着冠层光合持续期长短及其与光温资源的合理匹配,最终决定品种产量潜力是否能充分发挥。以往的作物模型研究结果^[50,52]为定量评价这些因子对提高超级稻产量潜力的影响打下了很好的基础。

综上所述,作者建议从叶片光合生理(光合电子传递、Rubisco 酶含量与活性、气孔与叶肉导度、叶绿体发育状况、磷酸丙糖运转等)、冠层光氮匹配(冠层光氮分布、氮素代谢、激素调控等)和根-冠水分平衡(根茎叶木质部发育、根系导水率、水通道蛋白表达等)这几个方面开展下一步研究,并在此基础上利用模型分析方法为辅助,揭示现代超级稻品种实现产量潜力的关键制约因素及其生理调控基础,并提出相应的栽培调控手段。

参考文献

- [1] 央广网.农业部通报:“超级稻”亩产已过千公斤[EB/OL]. http://china.cnr.cn/NewsFeeds/201410/t20141010_516575617.shtml.
- [2] Yang J, Zhang J. Grain-filling problem in ‘super’ rice[J]. *J Exp Bot*, 2010, 61: 1–5.
- [3] Gebbing T, Schnyder H. Pre-anthesis reserve utilization for protein and carbohydrate synthesis in grains of wheat[J]. *Plant Physiol*, 1999, 121: 871–878.
- [4] Takai T, Fukuta Y, Shirawa T, et al. Time-related mapping of quantitative trait loci controlling grain-filling in rice [J]. *J Exp Bot*, 2005, 56: 2 107–2 118.
- [5] Farquhar G D, von Caemmerer S, Berry J A. A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃ species [J]. *Planta*, 1980, 149: 78–90.
- [6] von Caemmerer S, Farquhar G, Berry J. Biochemical model of C₃ photosynthesis[C] // Laisk A, Nedbal L, Govindjee G, et al. Photosynthesis in silico: Understanding complexity from molecules to ecosystems. Dordrecht, The Netherlands: Springer, 2009: 209–230.
- [7] Kumar A, Li C, Portis Jr A R. Arabidopsis thaliana expressing a thermostable chimeric Rubisco activase exhibits enhanced growth and higher rates of photosynthesis at moderately high temperatures [J]. *Photosynth Res*, 2009, 100: 143–153.
- [8] Raines C A. Increasing photosynthetic carbon assimilation in C₃ plants to improve crop yield: current and future strategies Photosynthesis[J]. *Plant physiol*, 2011, 155: 36–42.
- [9] Kebeish R, Niessen M, Thiruveedhi K, et al. Chloroplastic photorespiratory bypass increases photosynthesis and biomass production in *Arabidopsis thaliana*[J]. *Nat Biotechnol*, 2007, 25: 593–599.
- [10] Miyagawa Y, Tamoi M, Shigeoka S. Overexpression of a cyanobacterial fructose-1, 6-/sedoheptulose-1, 7-bisphosphatase in tobacco enhances photosynthesis and growth [J]. *Nat Biotechnol*, 2001, 19: 965–969.
- [11] Stitt M, Lunn J, Usadel B. Arabidopsis and primary photosynthetic metabolism—more than the icing on the cake [J]. *Plant J*, 2010, 61: 1 067–1 091.
- [12] Yamori W, Takahashi S, Makino A, et al. The roles of ATP synthase and the cytochrome b6/f complexes in limiting chloroplast electron transport and determining photosynthetic capacity [J]. *Plant Physiol*, 2011, 155: 956–962.
- [13] Lieman-Hurwitz J, Rachmilevitch S, Mittler R, et al. Enhanced photosynthesis and growth of transgenic plants that express *ictB*, a gene involved in HCO₃⁻ accumulation in cyanobacteria [J]. *Plant Biotechnol J*, 2003, 1: 43–50.
- [14] Flexas J, Ribas-Carbó M, Hanson D T, et al. Tobacco aquaporin NtAQP1 is involved in mesophyll conductance to CO₂ in vivo [J]. *Plant J*, 2006, 48: 427–439.
- [15] Sade N, Gebretsadik M, Seligmann R, et al. The role of tobacco Aquaporin1 in improving water use efficiency, hydraulic conductivity, and yield production under salt stress [J]. *Plant Physiol*, 2010, 152: 245–254.
- [16] von Caemmerer S, Quick W P, Furbank R T. The development of C₄ rice: current progress and future challenges [J]. *Science*, 2012, 336: 1 671–1 672.
- [17] Sheehy J E, Ferrer A B, Mitchell P L, et al. How the rice crop works and why it needs a new engine[C] // Sheehy J E, Mitchell P L, Hardy B, eds. Charting new pathways to C₄ rice, Los Baños, Philippines: International Rice Research Institute, 2008: 3–26.
- [18] Gu J, Yin X, Struik P C, et al. Using chromosome introgression lines to map quantitative trait loci for photosynthesis parameters in rice (*Oryza sativa* L.) leaves under drought and well watered field conditions[J]. *J Exp Bot*, 2012, 63: 455–469.
- [19] Gu J, Yin X, Stomph T J, et al. Physiological basis of genetic variation in leaf photosynthesis among rice (*Oryza sativa* L.) introgression lines under drought and well-watered conditions[J]. *J Exp Bot*, 2012, 63: 5 137–5 153.
- [20] Adachi S, Baptista L Z, Sueyoshi T, et al. Introgression of two chromosome regions for leaf photosynthesis from an indica rice into the genetic background of a japonica rice[J]. *J Exp Bot*, 2014, 65: 2 049–2 056.
- [21] Adachi S, Tsuru Y, Nito N, et al. Identification and characterization of genomic regions on chromosomes 4 and 8 that control the rate of

- photosynthesis in rice leaves[J]. *J Exp Bot*, 2011, 62: 1927–1938.
- [22] Tayalaran R D, Adachi S, Ookawa T, et al. Hydraulic conductance as well as nitrogen accumulation plays a role in the higher rate of leaf photosynthesis of the most productive variety of rice in Japan [J]. *J Exp Bot*, 2011, 62: 4 067–4 077.
- [23] Yang X, Li Y, Ren B, et al. Drought-induced root aerenchyma formation restricts water uptake in rice seedlings supplied with nitrate[J]. *Plant Cell Physiol*, 2012, 53: 495–504.
- [24] Long S P, Zhu X, Naidu S L, et al. Can improvement in photosynthesis increase crop yields? [J]. *Plant, Cell and Environ*, 2006, 29: 315–330.
- [25] Zhu X G, Song Q, Ort D R. Elements of a dynamic systems model of canopy photosynthesis [J]. *Curr Opin Plant Biol*, 2012, 15: 237–244.
- [26] Evans J R. Improving photosynthesis [J]. *Plant Physiol*, 2013, 162: 1 780–1 793.
- [27] Song Q, Zhang G, Zhu X G. Optimal crop canopy architecture to maximise canopy photosynthetic CO₂ uptake under elevated CO₂—a theoretical study using a mechanistic model of canopy photosynthesis [J]. *Funct Plant Biol*, 2013, 40: 108–124.
- [28] Duvick D N. Genetic rates of gain in hybrid maize yields during the past 40 years[J]. *Maydica*, 1977, 22: 187–196.
- [29] Peng S, Khush G S, Virk P, et al. Progress in ideotype breeding to increase rice yield potential[J]. *Field Crop Res*, 2008, 108: 32–38.
- [30] Grindlay D J C. Towards an explanation of crop nitrogen demand based on the optimization of leaf nitrogen per unit leaf area [J]. *Agroecol Sci*, 1997, 128: 377–396.
- [31] Dreccer M F, Slafer G A, Rabbinge R. Optimization of vertical distribution of canopy nitrogen: an alternative trait to increase yield potential in winter cereals[J]. *J Crop Prod*, 1998, 1: 47–77.
- [32] Hikosaka K. Leaf canopy as a dynamic system: ecophysiology and optimality in leaf turnover[J]. *Ann Bot*, 2005, 95: 521–533.
- [33] Hirose T, Werger M J A. Maximizing daily canopy photosynthesis with respect to the leaf nitrogen allocation pattern in the canopy[J]. *Oecologia*, 1987, 72: 520–526.
- [34] Hikosaka K, Terashima I, Katoh S. Effects of leaf age, nitrogen nutrition and photon flux density on the distribution of nitrogen among leaves of a vine (*Ipomoea tricolor* Cav.) grown horizontally to avoid mutual shading of leaves[J]. *Oecologia*, 1994, 97: 451–457.
- [35] Drouet J L, Bonhomme R. Do variations in local leaf irradiance explain changes to leaf nitrogen within row maize canopies? [J]. *Ann Bot*, 1999, 84: 61–69.
- [36] Pons T L, Schieving F, Hirose T, et al. Optimization of leaf nitrogen allocation for canopy photosynthesis in *Lysimachia vulgaris* [C]//H Lambers, M L Cambridge, H Konings, et al. Causes and consequences of variation in growth rate and productivity of higher plants. The Hague, *The Netherlands: SPB Academic*, 1989: 175–186.
- [37] Anten N, Schieving F, Werger M. Patterns of light and nitrogen distribution in relation to whole canopy carbon gain in C₃ and C₄ mono- and dicotyledonous species[J]. *Oecologia*, 1995, 101: 504–513.
- [38] Yin X, Lantinga E A, Schapendonk A H C M, et al. Some quantitative relationships between leaf area index and canopy nitrogen content and distribution[J]. *Ann Bot*, 2003, 91: 893–903.
- [39] Hikosaka K. Optimal nitrogen distribution within a leaf canopy under direct and diffuse light [J]. *Plant Cell Environ*, 2014, 37: 2 077 – 2 085.
- [40] Dingkuhn M, Penning de Vries F W T, De Datta S K, et al. Concepts for a new plant type for direct seeded flooded tropical rice [C]// International Rice Research Institute. Direct seeded flooded rice in the Tropics. Los Baños, Philippines, 1991: 17–38.
- [41] Rousseaux M, Hall A, Sánchez R. Light environment, nitrogen content, and carbon balance of basal leaves of sunflower canopies [J]. *Crop Sci*, 1999, 39: 1 093–1 100.
- [42] Boonman A, Prinsen E, Gilmer F, et al. Cytokinin import rate as a signal for photosynthetic acclimation to canopy light gradients [J]. *Plant Physiol*, 2007, 143: 1 841–1 852.
- [43] Grindlay D J C, Sylvester-Bradley R, Scott R K. The relationship between canopy green area and nitrogen in the shoot[J] //G Lemaire, I G Burns, eds. Diagnostic Procedures for Crop N Management. Poitiers, France: INRA, 1995, 82: 53–60.
- [44] Lötscher M, Stroh K, Schnyder H. Vertical leaf nitrogen distribution in relation to nitrogen status in grassland plants [J]. *Ann Bot*, 2003, 92: 679–688.
- [45] Dreccer M F, Van Oijen M, Schapendonk A H C M, et al. Dynamics of vertical leaf nitrogen distribution in a vegetative wheat canopy: impact on canopy photosynthesis[J]. *Ann Bot*, 2000, 86: 821–831.
- [46] Sinclair T R, Shiraiwa T. Soybean radiation-use efficiency as influenced by nonuniform specific leaf nitrogen distribution and diffuse radiation [J]. *Crop Sci*, 1993, 33: 808–812.
- [47] Milroy S P, Bange M P, Sadras V O. Profiles of leaf nitrogen and light in reproductive canopies of cotton (*Gossypium hirsutum*) [J]. *Ann Bot*, 2001, 87: 325–333.
- [48] Prieto J A, Louarn G, Perez Pena J, et al. A leaf gas exchange model that accounts for intra - canopy variability by considering leaf nitrogen content and local acclimation to radiation in grapevine (*Vitis vinifera* L.) [J]. *Plant Cell Environ*, 2012, 35: 1 313–1 328.
- [49] Chen T W, Henke M, de Visser P H, et al. What is the most prominent factor limiting photosynthesis in different layers of a greenhouse cucumber canopy? [J]. *Ann Bot*, 2014, 114: 677–688.
- [50] Gu J, Yin X, Stomph T J, et al. Can exploiting natural genetic variation in leaf photosynthesis contribute to increasing rice productivity? A simulation analysis[J]. *Plant Cell Environ*, 2014, 37: 22–34.
- [51] Richards R A. Selectable traits to increase crop photosynthesis and yield of grain crops[J]. *J Exp Bot*, 2000, 51: 447–458.
- [52] Gu J, Yin X, Zhang C, et al. Linking ecophysiological modelling with quantitative genetics to support marker-assisted crop design for improved yields of rice (*Oryza sativa*) under drought stress[J]. *Ann Bot*, 2014, 114: 499–511.

Approaches to Improve Yield Potential of Super-rice from a Crop Physiological Perspective

GU Junfei, CHEN Ying, MAO Yiqi

(Jiangsu Key Laboratory of Crop Genetics and Physiology/Co-Innovation Center for Modern Production Technology of Grain Crops, Yangzhou University, Yangzhou, Jiangsu 225009, China; 1st author: gujf@yzu.edu.cn)

Abstract: Rice yield production is limited by the carbohydrate supply during grain-filling, which is unable to fill the large number of florets of rice plants, especially in the newly bred super-rice with numerous spikelets in a panicle. During grain-filling stage, carbohydrate supply depends on carbon from two sources: current photosynthetic assimilates and pre-stored assimilates in culms and leaf sheaths of rice plants. It is necessary to conduct the ecophysiological study on the sources of carbohydrate supply, which would enhance our understanding of limitations to yield potential in modern super-rice. The author summarized recent progresses in this field, and proposed that yield potential of modern ‘super’ rice could be improved by exploring ecophysiological properties of leaf photosynthesis, interaction of light and nitrogen distribution within canopy, and the relationship between root water uptake and leaf water potential. In the end, the author emphasized the role of modelling in integrating crop physiological knowledge to find the limitations to realizing super-rice yield potential and its physiological basis.

Key words: photosynthesis; yield potential; light and nitrogen distribution within canopy; relationship between root water uptake and leaf water potential

·综合信息·

广东省 2017 年审定通过的水稻新品种(上)

审定编号 (粤审稻)	品种名称	类型	选育单位	品种来源	区试产量 (kg/667m ²)	生试产量 (kg/667m ²)	全生育期 (d)
20170001	禅山占	籼型常规稻	广东省佛山市农业科学研究所	五山丝苗 / 小粒香	451.03	453.00	126~128
20170002	凤新丝苗	籼型常规稻	广东省东莞市中堂凤冲水稻科研站	凤枣丝苗 / 凤香丝苗	460.16	489.29	125~128
20170003	合莉油占	籼型常规稻	广东省农业科学院水稻研究所	玉香油占 / 合丝占 // 黄莉占	451.62	476.47	124~128
20170004	粤泰油占	籼型常规稻	广东省农业科学院水稻研究所	粤油丝苗 // 粤丰占 / GD9501 // 粤泰丝苗 // 粤华丝苗	447.87	481.95	127~129
20170005	田禾 1 号	籼型常规稻	广东省佛山市农业科学研究所	莉丰占 / 矮秀占 // 五山丝苗	437.25	452.64	126~130
20170006	五源占	籼型常规稻	广州市农业科学研究院、广州乾农农业科技发展有限公司	五山丝苗 / 桂农占 // 五山丝苗 / 广源占 5 号	449.60	473.20	126~128
20170007	南秀软占	籼型常规稻	广东省农业科学院水稻研究所	黄软占 / 黄秀占	445.42	456.76	125~127
20170008	双黄丝苗	籼型常规稻	广东省农业科学院水稻研究所	黄丰占 / 黄广占	448.12	461.79	125~127
20170009	华航 52 号	籼型常规稻	国家植物航天育种工程技术研究中心(华南农业大学)	华航 31 号 / 金农丝苗 南农业大学	442.11	471.84	126~130
20170010	山软 8 号	籼型常规稻	广东省佛山市农业科学研究所	丰富占 / 粤泰丝苗	435.41	444.95	127~128
20170011	绿银占	籼型常规稻	深圳隆平金谷种业有限公司	黄银占 / 丰粤占	450.92	483.74	127~129
20170012	固金占	籼型常规稻	广东省农业科学院水稻研究所	固广占 / 金丝软占	428.05	454.11	127~131
20170013	广晶软占	籼型常规稻	广东省农业科学院水稻研究所	黄广油占 // 玉晶占 / 合丰占	425.31	436.10	125~129
20170014	裕优占	籼型三系杂交稻	广东兆华种业有限公司	裕 A × 新香占	485.99	499.77	123~126
20170015	裕优锋占	籼型三系杂交稻	广东鲜美种苗发展有限公司	裕 A × 青丰占	489.68	501.03	118~121
20170016	炳优华占	籼型三系杂交稻	江西先农种业有限公司、中国水稻研究所、湖南杂交水稻研究中心	炳 1A × 华占	477.59	495.36	123~124
20170017	隆优 3155	籼型三系杂交稻	湖南隆平种业有限公司	隆香 634A × R3155	491.25	518.72	121~123
20170018	隆香优华占	籼型三系杂交稻	湖南隆平种业有限公司、中国水稻研究所、袁隆平农业高科股份有限公司	隆香 634A × 华占	506.83	543.00	122~123
20170019	星优 622	籼型三系杂交稻	广东源泰农业科技有限公司	星 A × 泰 622	481.06	501.69	123~125
20170020	吉田优 16	籼型三系杂交稻	广东天弘种业有限公司、广东省连山壮族瑶族自治县农业科学研究所	吉田 A × 弘恢 16	495.17	504.78	123~124
20170021	隆晶优华占	籼型三系杂交稻	湖南隆平高科种业科学研究院有限公司	隆晶 4302A × 华占	497.13	523.26	124~125

(下转第 12 页)